



## Diet-derived Flavonoids: Bridging Epidemiological Chemoprevention and Preclinical Anti-tumor Mechanisms in Clinical Oncology

## Neil B. Panchal<sup>1\*</sup> and Vipul M. Vaghela<sup>2</sup>

<sup>1</sup>Department of Pharmacy, Sumandeep Vidyapeeth Deemed to be University, Piparia, Waghodia, Vadodara - 391760, Gujarat, India; nbp9171@gmail.com <sup>2</sup>Department of Pharmaceutical Chemistry, A. R. College of Pharmacy and G. H. Patel Institute of Pharmacy, Vallabh Vidyanagar, Anand - 388120, Gujarat, India

## Abstract

Flavonoids are an abundantly consumed group of dietary polyphenols present in fruits, vegetables, teas, herbs and other plant-derived foods composed of a diphenylpropane (C6-C3-C6) ring structure, allowing subclassification into flavonols, flavones, flavan-3-ols, anthocyanins and isoflavones based on substitutions on the heterocyclic C ring. Multiple case-control studies and prospective cohort analyses reveal higher intake of certain flavonoid subgroups associated with reduced risk of various epithelial cancers like lung, breast, pancreatic, oral and liver. *In vitro* studies across diverse human cancer cell lines and in vivo, animal models demonstrate anticancer effects of select flavonoids either directly or in synergy with chemotherapy by targeting hallmark capabilities that enable tumours including resisting cell death, sustaining proliferation, inducing angiogenesis, activating invasion and metastasis. The well-explored anticancer mechanisms range from direct antioxidant activity, quenching free radicals and bolstering endogenous defenses; to anti-inflammatory signalling via NF-κB and cytokine modulation; epigenetic alterations by chromatin remodeling; to direct regulation of cell cycle controllers (CDKs, cyclins) and apoptotic mediators (caspases, Bcl-2. Early human trials mostly indicate the safe use of certain flavonoids and subclasses at tested doses however, progression to therapeutic benefit faces challenges like suboptimal systemic availability upon metabolism, unclear metabolite activities and study design limitations regarding delivery methods, combination treatments and clinical priority. In essence, dietary flavonoids exhibit pleiotropic pharmacological strengths against cancer progression warranting expanded translational research and human trials to develop formulations/delivery systems and validate targeted clinical integration, especially alongside chemotherapy regimens.

Keywords: Anticancer Activity, Epidemiological, Flavonoids, Molecular Mechanisms, Translational Research

## 1. Introduction

Flavonoids are a large group of polyphenolic compounds found ubiquitously in plants. Chemically, flavonoids consist of two aromatic rings linked together by a 3-carbon bridge, forming a common diphenyl propane skeleton<sup>1,2</sup>. Over 9,000 varietals of flavonoids have been identified and categorised into subclasses based on the oxidation state of the central pyran ring. The six major subclasses are flavonols, flavones, flavanones, flavan-3ols, anthocyanins, and isoflavones. Flavonoids differ in their distribution - some are found in all plant products (e.g. quercetin), while others are specific to particular foods (e.g. flavanones in citrus fruit)<sup>3</sup>.

Based on the oxidation state of ring C, flavonoids can be further divided into different subclasses<sup>4</sup> (Figure 1).

- Flavonols: Contain a 3-hydroxyflavone backbone with a ketone group on ring C (e.g. quercetin, kaempferol). They are found in fruits, vegetables, tea, and red wine<sup>5</sup>.
- Flavones: Contain an unsubstituted flavone backbone without hydroxylation on ring C (e.g. apigenin, luteolin). Found in parsley, celery, and chamomile tea<sup>6</sup>.

<sup>\*</sup>Author for correspondence

1634 Diet-derived Flavonoids: Bridging Epidemiological Chemoprevention and Preclinical Anti-tumor...





- Isoflavones: Contain the 3-phenylchromen-4-one backbone (e.g. daidzein, genistein). Largely found in soy and other legumes<sup>7</sup>.
- Flavanones: Contain a 2,3-dihydroflavone backbone with a saturated heterocycle. Found abundantly in citrus fruits<sup>8</sup>.
- Flavan-3-ols: Have a 2-phenyl-3,4-dihydro-2Hchromene skeleton (e.g. catechins). Found in tea, cocoa, black grapes and wine<sup>9</sup>.
- Anthocyanidins: Derived from the flavylium cation and are pigments in blue/ reddish fruits and vegetables<sup>10</sup>.

This classification is based on the structural differences in ring C, while substitutions to rings A and B contribute to the diversity among the over 9,000 identified flavonoids. Hydroxylation, methoxylation, prenylation, glycosylation and polymerisation further

add to the structural heterogeneity<sup>11</sup>. For example, the flavonol quercetin, abundantly found in onions, apples and broccoli, contains five hydroxyl groups, whereas kaempferol only has four<sup>12</sup>.

Dietary intake studies estimate the average person consumes about 1 gram of mixed flavonoids daily. The richest food sources are fruits, vegetables (especially onions, parsley, broccoli), tea, red wine and cocoa<sup>13</sup>. Herbs, spices, legumes and grains also contribute to intake. Though there is variation between regions and individual diets, flavonoid-rich items are common worldwide. The estimated per capita flavonoid intake in the United States is ~650 mg/day<sup>14</sup>.

For centuries, flavonoids have been used in botanical medicines worldwide. The first scientific report on flavonoids was in 1930 when Szent-Gyorgyi isolated citrin from lemon peel<sup>15</sup>. In the 1930s flavones were shown to reduce capillary bleeding. The term 'vitamin P', for permeability vitamin, was introduced - reflecting the vascular protective benefits observed. Since then thousands of studies have evaluated flavonoids. elucidating anti-inflammatory, antidiabetic, cardio-protective and anti-cancer effects<sup>3</sup>. Due to their abundance in the diet and evidence across epidemiological as well as clinical studies, regulatory agencies have recognized some flavonoids as Generally Recognized as Safe (GRAS). Wellestablished members include quercetin, catechins and soy isoflavones<sup>16</sup>.

The pleiotropic effects of flavonoids on health can be attributed to their molecular mechanisms of action. Being polyphenols, flavonoids exhibit excellent antioxidant capacity intracellularly and *in vitro*<sup>17</sup>. However newer evidence points to their interactions with cell signalling pathways that control inflammation, angiogenesis, cell proliferation, death and differentiation<sup>3</sup>. They modulate pathways by altering transcription factors and binding sites on receptors as well as affect downstream kinases like MAPK<sup>18</sup>. Recent work also reveals their epigenetic effects via chromatin remodelling and access to promoter regions<sup>19</sup>.

In conclusion, dietary flavonoids are an intensively studied group of plant polyphenols present in commonly consumed fruits, vegetables and plantderived foods and beverages. Their health protective effects span anti-inflammatory, anti-diabetic, anti-viral and anti-cancer benefits. This makes understanding their pharmacological mechanisms valuable for nutrition policies and therapeutic development.

## 2. Flavonoids and Cancer Prevention

Numerous epidemiological research has shown a negative correlation between the consumption of flavonoids found in fruits, vegetables, tea, and other sources and the risk of developing cancer.

Meta-analysis of case-control studies from China showed total flavonoid intake associated with reduced gastric cancer risk. The highest vs lowest analysis showed an odds ratio of 0.50 (95% CI 0.40–0.61). Epigallocatechin gallate from green tea is linked to a 31% decreased risk<sup>20,21</sup>.

European Prospective Investigation into Cancer and Nutrition examined total flavonoid subclass intake and liver cancer. The hazard ratio was 0.75 (95% CI 0.62– 0.91, Ptrend = 0.004) for hepatocellular carcinoma and 0.51 for flavonol subclass when comparing top vs bottom quintiles<sup>22</sup>.

A meta-analysis of 11 cohort studies revealed that breast cancer risk was lower for those in the highest versus the lowest quantile of flavonoid intake. Stratification by menopausal status showed stronger associations in postmenopausal women<sup>23,24</sup>.

A meta-analysis by Grosso *et al.* investigating dietary flavonoids for the risk of oral-pharynx and larynx cancers suggested a significant risk reduction. The odds ratio for total flavonoid intake was 0.60 and 0.47 for flavan-3-ol monomers<sup>25</sup>.

## 3. Proposed Mechanisms of Cancer Prevention

Multiple underlying mechanisms have been proposed that mediate the chemoprotective nature of flavonoids based on *in vitro* and animal data.

## 3.1 Antioxidant Activity

The phenolic structure of flavonoids enables them to act as hydrogen donors, metal ion chelators and free radical scavengers. They augment and interact with endogenous redox networks in cells.

Quercetin displays inhibition against peroxyl radicals, superoxide radicals and hydroxyl radicals. The

ortho-dihydroxy phenolic structure is crucial for this broad scavenging capacity<sup>26</sup>.

Myricetin, containing catechol and pyrogallol moieties, effectively inhibits DNA oxidation by Fenton reagents. It demonstrates cytoprotection in tert-butyl hydroperoxide models via free radical trapping<sup>27</sup>.

Fisetin increases the activities of glutathione peroxidase, glutathione reductase, catalase, and Nrf2 (Nuclear factor erythroid 2–related factor 2) signalling and suppresses nitric oxide generation in murine B16F10 melanoma cells treated with  $H_2O_2$  oxidative stress inducer. This reveals its role in fortifying cellular antioxidant defences<sup>28,29</sup>.

Benefits against DNA damage, lipid peroxidation and mitochondrial dysfunction reveal their ability to modulate cellular oxidative stress that drives carcinogenesis pathways.

#### 3.2 Anti-inflammatory Effects

Inflammation is now recognized as a key enabler of tumour growth, promoting cell proliferation, survival, invasion and angiogenesis.

Myricetin suppresses LPS-induced inflammatory activation in macrophages by downregulation of TNF $\alpha$ , IL-1 $\beta$ , IL-6 and COX2 expression by regulating the NF- $\kappa$ B and MAPK/ERK pathways<sup>30</sup>.

Similarly, apigenin inhibited LPS-stimulated TNF $\alpha$  production in murine peritoneal macrophages through suppression of NF- $\kappa$ B transcriptional activation and protein expression<sup>31,32</sup>.

In dextran sulfate sodium-induced colitis mice models, oral supplementation of quercetin, apigenin and luteolin demonstrated protective effects by lowering myeloperoxidase activity and preserving antioxidant defences<sup>33,34</sup>.

### 3.3 Modulation of Cell Signalling Pathways

Flavonoids can modulate mutagenic pathways operating in cancer via the regulation of a multitude of enzymes, transcription factors, receptors, cell cycle and death proteins.

Soy isoflavones like genistein suppress protein tyrosine kinases involved in phosphorylation of cell growth factor receptors and thereby regulate signalling pathways that control cell proliferation and differentiation<sup>35</sup>.

Silibinin, a flavonoid in milk thistle, down-regulates COX-2, iNOS expression, and associated cytokine signalling by inhibiting the phosphorylation of MAP kinases like p38 and JNK in skin cancer models<sup>36</sup>.

Quercetin inhibits the PI3K/Akt cascade by modulating the activation of its upstream regulators like HER2/3 and EGFR. This leads to reduced viability and migration in ovarian and cervical cancer models<sup>37,38</sup>.

### 3.4 Epigenetic Effects

Flavonoids can modify gene expression through their impact on epigenetic processes, including DNA methylation, alterations in histone structure, and the arrangement of nucleosomes.

Green tea polyphenols have been shown to reactivate methylation-silenced tumour suppressor genes like p16INK4a, retinoic acid receptor  $\beta$  (RAR $\beta$ ), O6-methylguanine-DNA methyltransferase (MGMT), human mutL homolog 1 (hMLH1), and glutathione S-transferase pi 1 (GSTP1) in skin, oesophageal, and prostate cancer cells<sup>39</sup>.

Apigenin triggers the halt of the cell cycle in human prostate cancer by causing acetylation of histone H3 and H4, which in turn activates the transcription of the p21 and p53 genes<sup>40,41</sup>.

#### 3.5 Induction of Apoptosis

Abrogation of apoptotic pathways is a key hallmark of malignant cells.

Silymarin causes the collapse of mitochondrial membrane potential, enhances caspase-3 activity, and downregulates Bcl-2, and Bcl-XL proteins while upregulating Bax expression to drive the intrinsic apoptotic pathway in human prostate, breast and bladder cancer cells<sup>42,43</sup>.

Similarly, genistein demonstrates caspasedependent apoptosis induction in MDA-MB-231 cells by suppressing Bcl-2 and Bcl-xL and activating Bax<sup>44,45</sup>.

Fisetin selective cytotoxicity against uveal melanoma cells involves the activation of ROS-dependent mitogen-activated protein kinases triggering the caspase cascade<sup>46-48</sup>.

Apigenin reactivates the TRAIL-R2 apoptotic pathway by upregulating death receptor expression in TRAIL-resistant prostate cancer cells<sup>49–52</sup>.

Inhibition of angiogenesis and metastasis by interfering with VEGF-stimulated endothelial cell proliferation, migration and proteolytic enzyme activity, flavonoids can inhibit tumour angiogenesis.

Methylated flavonoids like chrysin, tamarixetin, and silibinin show anti-angiogenic properties in rodent models by targeting the VEGF/bFGF-regulated ERK /FAK/MMP pathways. This controls endothelial cell migration, invasion and tube formation<sup>53</sup>.

Similarly, naringenin, hesperetin and nomilin among citrus flavonoids have revealed anti-angiogenic effects by downregulating VEGF, hypoxia-inducible factor 1-alpha, CD31, and vimentin expression in treated animals<sup>54–56</sup>.

Epigallocatechin gallate competitively inhibits the binding of VEGF165 to the NP1 receptor and disrupts downstream signalling by AKT and ERK1/2 associated with angiogenesis<sup>57,58</sup>.

Genistein also demonstrates anti-metastatic potential by downregulating matrix metalloproteinases and upregulating tissue inhibitors of metalloproteinases. Reduced cell motility and invasion are also achieved by modulation of genes like nm-23 and Kai1 that suppress metastasis<sup>59,60</sup>.

## 4. Anti-cancer Effects of Flavonoids

*In vitro* evidence, multiple *in vitro* studies reveal the potential anti-cancer effects of different flavonoid compounds across various human cancer cell lines:

Quercetin displays anti-proliferative activity in oral (KB, CAL-27), leukaemia (HL60, HEL, K562), gastric, colon, prostate, lung, bladder, ovarian and breast (MCF7, MDA-MB231) cancer cells. It causes cell cycle arrest and apoptosis induction across models<sup>61</sup>.

Similarly, apigenin-induced apoptosis and cell cycle arrest in pancreatic (PANC-1), prostate (PC3, 22Rv1), thyroid (BCPAP, TPC-1), cervical (HeLa, C33A) and haematological cancer cells through modulation of Bax/Bcl-2 genes and other proteins governing cell cycle progression<sup>62</sup>.

Genistein from soy demonstrated dose-dependent cytotoxic effects in cultures of the prostate (LNCaP, DU145), gastric (MGC-803), pancreatic (MiaPaCa-2), colorectal (HCT116), metastatic brain (F98) and ovarian (SKOV3) cancer cells via regulation of apoptosis, cell growth and invasion related signalling cascades among other pathways<sup>45,63,64</sup>.

## 5. Influence on Cancer Hallmarks

The anti-cancer mechanisms elucidated span across the hallmark capabilities tumour cells acquire:

## 5.1 Cell Cycle Arrest

Many studies reveal flavonoids like luteolin, quercetin and apigenin can induce cell cycle arrest at G0/G1 or G2/M phase by acting on cyclins, CDKs and their inhibitors like p21 and p27. Targets include Akt, FOXO and p53 pathways. The review discusses mechanisms like suppression of CDK1, CDK2, CDK4, CDK6, cyclin D1 and cyclin E1 expression in prostate, breast, cervical and liver cancer cells treated with various flavonoids. Downregulation of these cell cycle controllers causes arrest at growth phases<sup>65,66</sup>.

## 5.2 Proliferation and Growth Inhibition

Flavonoids suppress proliferation signals via PI3K/ Akt, JAK/STAT, Wnt and MAP kinase pathways. Genistein and EGCG prevent the activation of growth factor receptors like EGFR, ERBB2 and downstream routes. Silibinin causes G1 arrest by hindering hyperphosphorylation of retinoblastoma protein<sup>67</sup>.

## 5.3 Induction of Apoptosis

Quercetin intercalates in DNA strands triggering caspase cascade<sup>68</sup>. Flavanolsenhance p53 transcriptional activity and loss of mitochondrial potential to prompt apoptosis via mechanisms including JNK, NF-κBand PI3K/AKT inhibition<sup>69,70</sup>. Isoflavones modulate Bcl-2 proteins, upregulate death receptors signalling like Fas/ FasL<sup>62</sup>.

## 5.4 Synergism with Chemotherapy

Co-administration of certain flavonoids has revealed synergistic activity with conventional chemotherapy drugs:

Silibinin boosts the effectiveness of cisplatin in models of lung and ovarian cancer by altering the signalling pathways that regulate cell growth and programmed cell death<sup>71,72</sup>.

Similarly, the combinatorial potential of quercetin has been reported with cisplatin in ovarian, hepatocellular and gastric cancers owing to complementary mechanistic regulation of apoptosis-related proteins, microRNA signalling and reactive oxygen species generation<sup>65</sup>.

### 6. Most Potent Flavonoid Subclasses

Among the subclasses, evidence points to isoflavones, flavonols, and flavan-3-ols as the most potent categories exhibiting anti-cancer benefits (Figure 2).

#### 6.1 Isoflavones

Genistein is among the most extensively studies isoflavones showing activity across diverse cancer cell lines - prostate, bladder, gastric, pancreatic, melanoma etc. Attributed to the inhibition of angiogenesis, metastasis, proliferation and induction of differentiation pathways<sup>63</sup>.

### 6.2 Flavonols

Quercetin has displayed effective and selective cytotoxic effects across vast malignancies – oral, breast, lung, pancreatic, hepatoma, leukaemia etc by targeting growth factor receptors. Myricetin similarly exhibits anticancer benefits<sup>68</sup>.

#### 6.3 Flavan-3-ols

Tea catechins like EGCG have revealed tumour inhibitory capacity especially for cancers of the digestive tract like gastric, colon etc. Targets matrix metalloproteinases, mitochondria-mediated apoptosis and VEGF pathways<sup>73,74</sup>.

## 7. Flavonoid Bioavailability and Metabolism

### 7.1 Absorption, Metabolism and Excretion

Flavonoid absorption primarily takes place in the small intestine, yet the amount that reaches the bloodstream





and target tissues is affected by their release from food matrices and their stability. Intestinal, and hepatic metabolism further modify structures before secretion back into the intestine or urinary system.

In intestinal absorption, most data suggest passive diffusion of lipophilic flavonoids like isoflavones across the intestinal wall. Structural transformations by lactase phlorizin hydrolase, cytosolic  $\beta$ -glucosidase and membrane-bound  $\beta$ -glucuronidase enzymes aid this permeation. An efflux ABC transporter MRP2 limits their epithelial retention. Circulating metabolites then undergo hepatic biotransformations<sup>75</sup>.

Colonic Metabolism Unabsorbed flavonoids pass into the colon where catabolism by resident microflora releases aglycones that may re-enterhepatic circulation through the portal vein or get eliminated via faecal excretion. Microbial enzymatic activities give rise to various phenolic acids. Faecal microbiome stability and compositions hence govern bioavailability to an extent<sup>76</sup>.

Hepatic metabolism and elimination pathways: methylation, glucuronidation and sulfation represent the key hepatic modifications as phase II conjugation reactions to facilitate biliary and urinary secretion: Catechol-o-methyl transferase (COMT) catalyses the O-methylation of dihydroxyl groups on ring scaffolds. UDP-glucuronosyltransferase adds glucuronic acid moieties, especially at positions with a hydroxyl group, like ring C or position 7 on ring A of flavonoids. Sulfotransferases transfer sulfate groups to hydroxyl structures into sulfo-conjugated forms like quercetin-3'-sulfate before urinary elimination<sup>77</sup>.

### 7.2 Factors Affecting Bioavailability

Flavonoid bioavailability exhibits extensive personal variations attributed to:

## 7.2.1 Dietary Source

Glycosides from onions display nearly 10 times greater bioavailability over other food matrices. Solubilizing effects of a protein-rich meal matrix can also enhance uptake than carbohydrate-rich meals<sup>78</sup>.

### 7.2.2 Supplement-drug Interactions

Synergistic effects on absorption have been noted for quercetin and genistein when co-administered with

fruit juice, tea or wine components like ellagic acid and EGCG possibly owing to regulated efflux transport<sup>79,80</sup>.

## 7.2.3 Gut Microbial Community

Intestinal bacteria mediate metabolite production from nonabsorbable forms and hence direct systemic exposure. Prenylated chrysin metabolites are unique to mice harbouring intestinal lactobacilli. Inter-individual variability in microbiome underlies response diversity<sup>81,82</sup>.

## 7.2.4 Gene Polymorphisms

Variations in metabolic or transporter enzymes like COMT, UGT and ABC efflux carriers due to single nucleotide changes alter metabolite ratios and kinetics substantially between subjects<sup>83,84</sup>. Quercetin bioavailability displays a nearly 10-fold range based on such genetic differences<sup>80</sup>.

## 8. Strategies for Enhanced Bioavailability

Emerging approaches targeting the ADME framework to promote circulating flavonoid levels include:

## 8.1 Synthesis of Analogues

Structural analogues of hesperetin, and naringenin created through methylation, halogenation, and sulfonylation exhibit enhanced metabolic stability and membrane permeation. 4'-Fluoro-naringenin exhibits a tenfold increase in bioavailability compared to the native compound<sup>85,86</sup>.

## 8.2 Advanced Delivery Systems

Lipid-based vesicles, nanoemulsions, and solid dispersions protect the integrity of polyphenols through GI transit while improving mucosal permeation. A quercetin nanoemulsion established nearly 20 times more absorption than unformulated control in rodent PK studies<sup>87–89</sup>.

## 8.3 Modification of Sites Prone to Metabolism

Blocking positions vulnerable to sulfation, glucuronidation can retain potency. Synthetic enrichment of 7-O-glycosides demonstrates resilience

against phase II losses while preserving antiproliferative effects<sup>90</sup>.

## 9. Pharmacokinetics of Flavonoid Subclasses: Absorption and Metabolism Differences

## 9.1 Flavanones

Glycosylated flavanones have superior systemic levels over aglycones. Hesperetin bioavailability increased 5 fold owing to sugar moieties or rutinosides<sup>91,92</sup>.

## 9.2 Flavonols

Quercetin glucosides get effectively hydrolyzed by LPH and cytosolic  $\beta$ -glucosidase before absorption into circulation mainly as glucuronidated conjugates. About 70% of ingested quercetin enters the blood in metabolized forms<sup>75</sup>.

## 9.3 Isoflavones

Genistein and daidzein show nearly 100% absorption with peak levels at 6-8 hours. However, about 75-85% circulate as glucuronides and sulfoglucuronides while only 5-20% retain active unconjugated structures due to extensive phase II metabolism<sup>93</sup>.

## 9.4 Anthocyanins

Rapid phase II modifications form methylated, glucuronidated and sulfated metabolites although at very low fractions (2-5 %) owing to limited intrinsic absorption. Proanthocyanidins on polymerization are not absorbed at all in native forms<sup>94</sup>.

In essence, the position, type and extent of substitutions govern the absorption, transformation and pharmacokinetic profiles of each flavonoid category following oral ingestion.

## 10. Human Clinical Trials for Flavonoids in Cancer

Both epidemiological observational studies, as well as controlled interventional trials, have been conducted to examine associations of flavonoid intake with cancer outcomes in humans.

Epidemiological Studies Multiple cohort analyses reveal beneficial correlations.

Meta-analysis of 21 population studies found high vs low analysis of total flavonoid consumption associated with significantly reduced risk for lung cancer (RR 0.81; 95% CI 0.75-0.88). The strongest reductions were shown for the flavonol and flavone subclasses<sup>95,96</sup>.

Among 53 studies examining carotenoids and flavonoids for pancreatic cancer, higher dietary anthocyanidins, flavan-3-ols, flavones and flavonols displayed risk reductions between 14-22% in dose-response analysis<sup>97,98</sup>.

Investigation into breast cancer recurrence among 1900+ survivors indicated >6 servings/week of fruits had a 28% lower likelihood than those with <3 servings/week<sup>99</sup>. A combination of fruit and vegetables, berries, and flavonoids specifically correlated with risk reductions in stratified models adjusting for confounders<sup>100</sup>.

However, such retrospective analyses have inherent biases in measurement, reporting and control for confounders that limit causal evidence. Controlled trials help address those limitations.

## **10.1 Interventional Studies**

A double-blind placebo-controlled study of prostate cancer patients with flavonoid supplements (600 mg/d catechins, 100 mg quercetin) revealed improvements in PSA levels, cholesterol profile without toxicity along with better functionality scores, suggesting chemopreventive benefits. Reductions were also seen in markers of inflammation, angiogenesis and apoptosis regulation<sup>101</sup>.

Among ovarian cancer patients, 14-day preoperative soy isoflavone supplementation (200 mg/d) indicated modulation of pathways associated with cell cycle regulation, DNA repair, and apoptosis compared to the placebo group in subsequent tumour analysis indicating mechanisms that likely mediate protective actions<sup>102</sup>.

Other studies reflect prevention potential - a phase I dose-escalation study in oral premalignant lesions indicated safe use of muscadine grape skin extract containing anthocyanins, and flavonols up to 1400 mg daily without systemic toxicity through 12 weeks while showing some histological improvements on biopsy<sup>103</sup>.

## **10.2 Challenges in Translational Research**

However, several challenges remain in translating preclinical knowledge into clinical therapeutic contexts:

Limited bioavailability: Intrinsic low oral absorption limits tissue exposures to exert optimal effects, especially at higher disease burden states in patients vs. preventative preclinical models<sup>104</sup>.

Complex metabolism: Flavonoid transformations by various tissue enzymes yield metabolites with unclear functional activities in humans. A better understanding of biologically active structures and routes is essential<sup>105</sup>.

Priorities for trials: Systemically delivered phase II/III clinical trials on formulated agents focusing on realistic tissue accumulation, pharmacodynamic biomarkers from preclinical models and clinically recommended combination regimens are lacking but are crucial to determining therapeutic index for highrisk populations<sup>106</sup>.

In essence, epidemiological data provides support while clinical studies have largely affirmed the safety of flavonoids at used doses. Advancing efficacy-focused investigation with innovative delivery systems in targeted trials remains vital to unlocking clinical promise and supporting clinical integration for cancer patients.

## 11. Safety and Side Effects of Flavonoids in Cancer Patients

Flavonoids, a diverse group of polyphenolic compounds found abundantly in various plant-based foods, have garnered significant attention for their potential pharmacological activity in cancer treatment. However, a thorough evaluation of the safety profile is imperative before considering their integration into cancer therapy. This section delves into the existing literature, presenting a meticulous analysis of the safety aspects of flavonoids in cancer patients, including potential side effects and interactions with conventional cancer therapies (Figure 3).

## 11.1 Safety Profile

Numerous studies have explored the safety of flavonoids in both preclinical and clinical settings. GRAS by regulatory bodies, flavonoids exhibit a favourable safety profile when consumed through dietary sources. However, caution is warranted when considering therapeutic doses in the context of cancer treatment.

## **11.2 Preclinical Safety Assessments**

Preclinical studies have provided valuable insights into the safety of flavonoids. While these compounds demonstrate low toxicity in normal cells, their effects on cancer cells are context-dependent. The diverse mechanisms by which flavonoids exert anticancer effects may also influence normal cellular functions<sup>107</sup>. Rigorous preclinical safety assessments are crucial for understanding potential off-target effects and establishing a therapeutic window.

## 11.3 Clinical Safety Data

Clinical trials investigating the safety of flavonoids in cancer patients have reported mixed findings. Most trials suggest a good tolerability profile, with minimal adverse effects observed at standard doses<sup>108</sup>. Frequently observed adverse effects encompass digestive issues (such as nausea and diarrhoea) and minor allergic responses. However, the incidence of adverse events varies across studies, emphasizing the need for standardized reporting and larger patient cohorts.

## **11.4 Possible Adverse Reaction** *11.4.1 Gastrointestinal Disturbances*

The most frequently reported side effects associated with flavonoid consumption in cancer patients are mild gastrointestinal disturbances. Nausea, abdominal discomfort, and diarrhoea have been reported in a subset of patients, but these effects are generally transient and reversible<sup>109</sup>.

### 11.4.2 Allergic Reactions

While rare, some individuals may experience allergic reactions to specific flavonoids. Skin rashes, itching, and, in severe cases<sup>110</sup>, anaphylaxis has been documented. Clinicians should be vigilant in monitoring patients for any signs of allergic responses, especially in those with a history of allergies.

#### 11.4.3 Hepatotoxicity

Limited cases of hepatotoxicity have been reported, prompting the need for careful monitoring of liver 1642 Diet-derived Flavonoids: Bridging Epidemiological Chemoprevention and Preclinical Anti-tumor...



Figure 3. Safety and side effects of flavonoids in cancer patients.

function in patients undergoing flavonoid-based therapies<sup>111</sup>. This is particularly relevant when considering long-term treatment or high-dose regimens.

# 12. Interactions with Conventional Cancer Therapies

## 12.1 Chemotherapeutic Agents

Interactions between flavonoids and conventional chemotherapeutic agents are complex and multifaceted. While some studies suggest synergistic effects that enhance the efficacy of chemotherapy, others indicate potential antagonism or interference with drug metabolism. Close monitoring of patients receiving concurrent flavonoid and chemotherapy regimens is essential to assess both therapeutic efficacy and the risk of adverse events<sup>112</sup>.

## 12.2 Antioxidant Effects and Radiation Therapy

Flavonoids are known for their antioxidant properties, which could theoretically interfere with the oxidative stress induced by radiation therapy. Studies exploring this interaction have yielded conflicting results, highlighting the need for further investigation to elucidate the impact of flavonoids on the efficacy of radiation therapy<sup>113</sup>.

## 13. Future Directions and Challenges

As we navigate the evolving landscape of cancer research and therapeutic interventions, the exploration

of flavonoids as potential agents for cancer treatment has unveiled promising avenues. However, a comprehensive understanding of the future directions and acknowledgement of current challenges and limitations is essential for steering this field towards impactful clinical applications (Figure 4).

## 13.1 Identification of Novel Flavonoids

The vast array of flavonoids in nature offers a rich source for potential therapeutic compounds. Future research should focus on the discovery and isolation of novel flavonoids with enhanced bioavailability, specificity to cancer cells, and potent anticancer activities<sup>114,115</sup>.

## **13.2 Precision Medicine Approaches**

The heterogeneity of cancer necessitates a personalized approach. Future studies should investigate the potential of flavonoids in precision medicine, considering individual variations in metabolism, genetic makeup, and the specific molecular characteristics of tumours<sup>116</sup>.

## 13.3 Combination Therapies

Investigating the synergistic effects of flavonoids with conventional cancer therapies holds promise. Combinatorial approaches could enhance therapeutic efficacy, reduce adverse effects, and potentially overcome resistance mechanisms that limit the effectiveness of current treatments<sup>117,118</sup>.

## 13.4 Understanding Mechanisms of Action

Unraveling the intricate molecular mechanisms underlying flavonoid-induced anticancer effects is crucial<sup>11</sup>. Future research should employ advanced technologies, such as single-cell omics,





to elucidate how flavonoids modulate signalling pathways, induce apoptosis, and impact the tumour microenvironment<sup>119</sup>.

#### 13.5 Exploration of Nanoformulations

Enhancing the bioavailability and stability of flavonoids through nanoformulations represents a promising avenue<sup>120</sup>. Nanoparticles can improve drug delivery, targeting, and release, potentially overcoming limitations related to flavonoid solubility and degradation<sup>121</sup>.

#### 13.6 Evaluation of Long-term Effects

Long-term safety and efficacy data for flavonoids in cancer patients are scarce<sup>122</sup>. Future studies should focus on conducting well-designed, longitudinal trials to assess the sustained effects of flavonoid interventions, ensuring a comprehensive understanding of their impact on cancer progression and patient outcomes<sup>123</sup>.

## 13.7 Integration of Preclinical and Clinical Data

Bridging the gap between preclinical and clinical research is imperative. Future studies should emphasize the translation of preclinical findings into meaningful clinical applications, addressing discrepancies and optimizing study designs to facilitate successful transitions from bench to bedside<sup>124–126</sup>.

## 14. Challenges and Limitations in Flavonoid Research for Cancer Treatment

#### 14.1 Bioavailability Issues

The limited bioavailability of certain flavonoids remains a significant challenge. Factors such as poor



Figure 5. Challenges and limitations of flavonoids research.

absorption, rapid metabolism, and limited distribution to target tissues hinder their therapeutic potential<sup>78,127</sup>. Overcoming these challenges requires innovative drug delivery strategies (Figure 5).

## 14.2 Lack of Standardization in Clinical Trials

Inconsistencies in the design and reporting of clinical trials hinder the comparison of results across studies<sup>128</sup>. Standardization of protocols, outcome measures, and reporting criteria is essential for generating reliable evidence and drawing meaningful conclusions<sup>129</sup>.

## 14.3 Inter-individual Variability

The response to flavonoid treatment varies among individuals due to genetic, dietary, and lifestyle factors<sup>130,131</sup>. Future research should explore ways to tailor flavonoid interventions to specific patient profiles, considering factors such as gut microbiota composition and genetic polymorphisms<sup>132</sup>.

## 14.4 Limited Understanding of Optimal Dosing

Determining the optimal dosage and duration of flavonoid treatment remains a challenge<sup>133</sup>. Future research should focus on establishing dose-response relationships, considering factors such as patient characteristics, cancer type, and stage<sup>134</sup>.

## 14.5 Complexity of Studying Combinatorial Therapies

Investigating the interactions between flavonoids and conventional cancer therapies presents methodological and logistical challenges<sup>135</sup>. Standardized methodologies and rigorous study designs are needed to assess the safety and efficacy of combination therapies accurately<sup>136</sup>.

### 14.6 Financial and Logistical Barriers

Conducting large-scale clinical trials and research initiatives requires significant financial and logistical resources<sup>137</sup>. Collaborative efforts between academia, industry, and regulatory agencies are essential to overcome these barriers and facilitate robust research endeavours<sup>138</sup>.

## 15. Conclusion

Flavonoids are an abundantly consumed group of dietary polyphenols present in fruits, vegetables,

teas, herbs and other plant-derived foods. Composed of a diphenylpropane (C6-C3-C6) ring structure, over 9,000 varietals exist classified as flavonols, flavones, flavan-3-ols, anthocyanins and isoflavones based on substitutions on the heterocyclic C ring. Extensive epidemiological evidence from casecontrol and prospective cohort studies reveals higher flavonoid subclass intake is associated with reduced risk of various epithelial cancers like lung, breast, pancreatic, oral and liver. Supporting in vitro analysis across diverse human cancer cell lines and in vivo animal models demonstrates direct and synergistic anticancer effects of certain flavonoids by targeting proliferation, cell cycle, apoptosis, metastasis and angiogenesis pathways. The wellelucidated mechanisms range from antioxidant activity, quenching free radicals and bolstering endogenous defences; to anti-inflammatory signalling via NF-kBand cytokine modulation; epigenetic alterations by chromatin remodelling; to direct regulation of cell cycle controllers (CDKs, cyclins) and apoptotic proteins (caspases, Bcl-2). Synergism with various chemotherapy drugs has been elucidated for enhanced anticancer efficacy. While early human trials mostly indicate safe use and potential chemopreventive efficacy, the realization of therapeutic benefit faces challenges like suboptimal systemic availability upon metabolism, unclear metabolite activities and study design limitations regarding delivery methods, combination treatments and clinical priority. In essence, dietary flavonoids exhibit multifaceted anticancer strengths warranting expanded translational research and human trials to develop formulations/delivery systems and validate targeted clinical integration.

## 16. Acknowledgement

We extend our deepest appreciation to Mr. Biren S. Panchal for his crucial guidance and unwavering support throughout this study.

## 17. References

1. Xiao ZP, Peng ZY, Peng MJ, Yan WB, Ouyang YZ, Zhu HL. Flavonoids health benefits and their molecular mechanism.

## 1646 Diet-derived Flavonoids: Bridging Epidemiological Chemoprevention and Preclinical Anti-tumor...

Mini Rev Med Chem. 2011; 11(2):169-77. https://doi. org/10.2174/138955711794519546 PMid:21222576

- Kumar S, Pandey AK. Chemistry and biological activities of flavonoids: An overview. Sci World J. 2013; 2013:16. https:// doi.org/10.1155/2013/162750 PMid:24470791 PMCid: PMC3891543
- Panche AN, Diwan AD, Chandra SR. Flavonoids: An overview. J Nutr Sci. 2016; 5:e47. https://doi.org/10.1017/ jns.2016.41 PMid:28620474 PMCid: PMC5465813
- Havsteen BH. The biochemistry and medical significance of the flavonoids. Pharmacol Ther. 2002; 96(2-3):67-202. https://doi.org/10.1016/S0163-7258(02)00298-X PMid: 12453566
- Mahmud AR, Ema TI, Siddiquee MFR, Shahriar A, Ahmed H, Mosfeq-Ul-Hasan M, *et al.* Natural flavonols: actions, mechanisms, and potential therapeutic utility for various diseases. Beni-Suef Univ J Basic Appl Sci. 2023; 12(1):1-18. https://doi.org/10.1186/s43088-023-00387-4 PMid:37216013 PMCid: PMC10183303
- Singh M, Kaur M, Silakari O. Flavones: An important scaffold for medicinal chemistry. Eur J Med Chem. 2014; 84:206-39. https://doi.org/10.1016/j.ejmech.2014.07.013 PMid:25019478
- Křížová L, Dadáková K, Kašparovská J, Kašparovský T. Isoflavones. Molecules. 2019; 24(6). https://doi.org/10.3390/ molecules24061076 PMid:30893792 PMCid: PMC6470817
- Shen N, Wang T, Gan Q, Liu S, Wang L, Jin B. Plant flavonoids: Classification, distribution, biosynthesis, and antioxidant activity. Food Chem. 2022; 383:132531.https:// doi.org/10.1016/j.foodchem.2022.132531 PMid:35413752
- Aron PM, Kennedy JA. Flavan-3-ols: nature, occurrence and biological activity. Mol Nutr Food Res. 2008; 52(1):79-104. https://doi.org/10.1002/mnfr.200700137 PMid:18081206
- Khoo HE, Azlan A, Tang ST, Lim SM. Anthocyanidins and anthocyanins: coloured pigments as food, pharmaceutical ingredients, and the potential health benefits. Food Nutr Res. 2017; 61(1). https://doi.org/10.1080/16546628.2017.13 61779 PMid:28970777 PMCid: PMC5613902
- Ramesh P, Jagadeesan R, Sekaran S, Dhanasekaran A, Vimalraj S. Flavonoids: Classification, function, and molecular mechanisms involved in bone remodelling. Front Endocrinol (Lausanne). 2021; 12. https://doi.org/10.3389/ fendo.2021.779638 PMid:34887836 PMCid: PMC8649804
- Dabeek WM, Marra MV. Dietary quercetin and kaempferol: Bioavailability and potential cardiovascular-related bioactivity in humans. Nutrients. 2019; 11(10). https:// doi.org/10.3390/nu11102288 PMid:31557798 PMCid: PMC6835347
- Pérez-Jiménez J, Neveu V, Vos F, Scalbert A. Identification of the 100 richest dietary sources of polyphenols: An application of the phenol-explorer database. Eur J Clin Nutr. 2010; 64(Suppl 3):S112-20. https://doi.org/10.1038/ ejcn.2010.221 PMid:21045839

- 14. Ock KC, Sang JC, Song WO. Estimated dietary flavonoid intake and major food sources of U.S. adults. J Nutr. 2007; 137(5):1244-52. https://doi.org/10.1093/jn/137.5.1244 PMid:17449588
- Seashore RH, Mccollom IN. The chemical nature of vitamin C. Science. 1932; 75:357-9. https://doi.org/10.1126/ science.75.1944.358 PMid:17750034
- CFR. CFR Code of Federal Regulations Title 21 (cited 2023 Nov 25). Available from: http://www.accessdata.fda. gov/scripts/cdrh/cfdocs/cfcfr/CFRSearch.cfm?fr=50.25
- Pietta PG. Flavonoids as antioxidants. J Nat Prod. 2000;
  63(7):1035-42. https://doi.org/10.1021/np9904509 PMid: 10924197
- Williams RJ, Spencer JPE, Rice-Evans C. Flavonoids: Antioxidants or signalling molecules? Free Radic Biol Med. 2004; 36(7):838-49. https://doi.org/10.1016/j. freeradbiomed.2004.01.001 PMid:15019969
- Hananya N, Koren S, Muir TW. Interrogating epigenetic mechanisms with chemically customized chromatin. Nat Rev Genet. 2023. p. 1-17. Available from: https://www. nature.com/articles/s41576-023-00664-z
- Bo Y, Sun J, Wang M, Ding J, Lu Q, Yuan L. Dietary flavonoid intake and the risk of digestive tract cancers: A systematic review and meta-analysis. Sci Rep. 2016; 6:24836. https:// doi.org/10.1038/srep24836 PMid:27112267 PMCid: PMC4845003
- Petrick JL, Steck SE, Bradshaw PT, Trivers KF, Abrahamson PE, Engel LS, *et al.* Dietary intake of flavonoids and oesophageal and gastric cancer: Incidence and survival in the United States of America (USA). Br J Cancer. 2015; 112(7):1291-300. https://doi.org/10.1038/bjc.2015.25 PMid:25668011 PMCid: PMC4385952
- 22. Zamora-Ros R, Agudo A, Luján-Barroso L, Romieu I, Ferrari P, Knaze V, *et al.* Dietary flavonoid and lignan intake and gastric adenocarcinoma risk in the European Prospective Investigation into Cancer and Nutrition (EPIC) study. Am J Clin Nutr. 2012; 96(6):1398-408. https://doi. org/10.3945/ajcn.112.037358 PMid:23076618
- Theodoratou E, Kyle J, Cetnarskyj R, Farrington SM, Tenesa A, Barnetson R, *et al.* Dietary flavonoids and the risk of colorectal cancer. Cancer Epidemiol Biomarkers Prev. 2007; 16(4):684-93. https://doi.org/10.1158/1055-9965.EPI-06-0785 PMid:17416758
- 24. Chang H, Lei L, Zhou Y, Ye F, Zhao G. Dietary flavonoids and the risk of colorectal cancer: An updated meta-analysis of epidemiological studies. Nutrients. 2018; 10(7). https:// doi.org/10.3390/nu10070950 PMid:30041489 PMCid: PMC6073812
- 25. Grosso G, Micek A, Godos J, Pajak A, Sciacca S, Galvano F, et al. Dietary flavonoid and lignan intake and mortality in prospective cohort studies: Systematic review and doseresponse meta-analysis. Am J Epidemiol. 2017; 185(12):1304-16. https://doi.org/10.1093/aje/kww207 PMid:28472215

- Russo M, Spagnuolo C, Tedesco I, Bilotto S, Russo GL. The flavonoid quercetin in disease prevention and therapy: facts and fancies. Biochem Pharmacol. 2012; 83(1):6-15. https:// doi.org/10.1016/j.bcp.2011.08.010 PMid:21856292
- Pietta PG, Gardana C, Mauri PL, Maffei-Facino R, Carini M. Identification of flavonoid metabolites after oral administration to rats of a *Ginkgo biloba* extract. J Chromatogr B Biomed Sci Appl. 1995; 673(1):75-80. https:// doi.org/10.1016/0378-4347(95)00252-E PMid: 8925077
- Naeimi AF, Alizadeh M. Antioxidant properties of the flavonoid fisetin: An updated review of *in vivo* and *in vitro* studies. Trends Food Sci Technol. 2017; 70:34-44. https:// doi.org/10.1016/j.tifs.2017.10.003
- 29. Ravula AR, Teegala SB, Kalakotla S, Pasangulapati JP, Perumal V, Boyina HK. Fisetin, a potential flavonoid with multifarious targets for treating neurological disorders: An updated review. Eur J Pharmacol. 2021; 910:174492. https://doi.org/10.1016/j.ejphar.2021.174492 PMid:34516952
- 30. Jang JH, Lee SH, Jung K, Yoo H, Park G. Inhibitory effects of myricetin on lipopolysaccharide-induced neuroinflammation. Brain Sci. 2020; 10(1):32. https://doi. org/10.3390/brainsci10010032 PMid:31935983 PMCid: PMC7016734
- Nicholas C, Batra S, Vargo MA, Voss OH, Gavrilin MA, Wewers MD, *et al.* Apigenin blocks lipopolysaccharideinduced lethality *in vivo* and proinflammatory cytokines expression by inactivating NF-kb through the suppression of p65 phosphorylation. J Immunol. 2007; 179(10):7121-7. https://doi.org/10.4049/jimmunol.179.10.7121 PMid: 17982104
- Ye X, Zhu M, Che X, Wang H, Liang XJ, Wu C, et al. Lipopolysaccharide induces neuroinflammation in microglia by activating the MTOR pathway and downregulating Vps34 to inhibit autophagosome formation. J Neuroinflammation. 2020; 17(1):1-17. https://doi.org/ 10.1186/s12974-019-1644-8 PMid:31926553 PMCid: PMC6954631
- 33. Chassaing B, Aitken JD, Malleshappa M, Vijay-Kumar M. Dextran Sulfate Sodium (DSS)-induced colitis in mice. Curr Protoc Immunol. 2014; 104(SUPPL.104). https:// doi.org/10.1002/0471142735.im1525s104 PMid:24510619 PMCid: PMC3980572
- 34. Park YH, Kim N, Shim YK, Choi YJ, Nam RH, Choi YJ, et al. Adequate dextran sodium sulfate-induced colitis model in mice and effective outcome measurement method. J Cancer Prev. 2015; 20(4):260. https://doi. org/10.15430/JCP.2015.20.4.260 PMid:26734588 PMCid: PMC4699753
- 35. Saeidnia S, Manayi A, Abdollahi M. From *in vitro* experiments to *in vivo* and clinical studies; pros and cons. Curr Drug Discov Technol. 2015; 12(4):218-24. https://doi. org/10.2174/1570163813666160114093140 PMid:26778084

- 36. Emadi SA, Rahbardar MG, Mehri S, Hosseinzadeh H. A review of therapeutic potentials of milk thistle (*Silybum marianum* L.) and its main constituent, silymarin, on cancer, and their related patents. Iran J Basic Med Sci. 2022; 25(10):1166.
- 37. Fahrenholtz CD, Swanner J, Ramirez-Perez M, Singh RN. Heterogeneous responses of ovarian cancer cells to silver nanoparticles as a single agent and in combination with cisplatin. J Nanomater. 2017; 2017. https://doi. org/10.1155/2017/5107485 PMid:30034459 PMCid: PMC6052800
- Yin M, Xu X, Han H, Dai J, Sun R, Yang L, *et al*. Preparation of triangular silver nanoparticles and their biological effects in the treatment of ovarian cancer. J Ovarian Res. 2022; 15(1):1-14. https://doi.org/10.1186/s13048-022-01056-3 PMid:36411490 PMCid: PMC9680130
- 39. Fang MZ, Wang Y, Ai N, Hou Z, Sun Y, Lu H, *et al.* Tea polyphenol (-)-Epigallocatechin-3-gallate inhibits DNA methyltransferase and reactivates methylation-silenced genes in cancer cell lines. Cancer Res. 2003; 63(22):7563-70.
- Shankar E, Goel A, Gupta K, Gupta S. Plant flavone apigenin: An emerging anticancer agent. Curr Pharmacol Reports. 2017; 3(6):423. https://doi.org/10.1007/s40495-017-0113-2 PMid:29399439 PMCid: PMC5791748
- Hnit SST, Yao M, Xie C, Bi L, Wong M, Liu T, *et al.* Apigenin impedes cell cycle progression at the G2 phase in prostate cancer cells. Discov Oncol. 2022; 13(1):44. https://doi. org/10.1007/s12672-022-00505-1 PMid:35670862 PMCid: PMC9174405
- 42. Deep G, Agarwal R. Antimetastatic efficacy of silibinin: molecular mechanisms and therapeutic potential against cancer. Cancer Metastasis Rev. 2010; 29(3):447-63. https:// doi.org/10.1007/s10555-010-9237-0 PMid:20714788 PMCid: PMC3928361
- 43. Agarwal C, Wadhwa R, Deep G, Biedermann D, Gažák R, Křen V, *et al.* Anti-cancer efficacy of silybin derivatives — a structure-activity relationship. PLoS One. 2013;8(3). https:// doi.org/10.1371/journal.pone.0060074 PMid:23555889 PMCid: PMC3610875
- 44. Li Z, Li J, Mo B, Hu C, Liu H, Qi H, et al. Genistein induces cell apoptosis in MDA-MB-231 breast cancer cells via the mitogen-activated protein kinase pathway. Toxicol Vitr. 2008; 22(7):1749-53. https://doi.org/10.1016/j. tiv.2008.08.001 PMid:18761399
- 45. Tuli HS, Tuorkey MJ, Thakral F, Sak K, Kumar M, Sharma AK, et al. Molecular mechanisms of action of genistein in cancer: Recent advances. Front Pharmacol. 2019; 10. https://doi.org/10.3389/fphar.2019.01336 PMid:31866857 PMCid: PMC6910185
- 46. Rahmani AH, Almatroudi A, Allemailem KS, Khan AA, Almatroodi SA. The potential role of fisetin, a flavonoid in cancer prevention and treatment. Molecules. 2022;

27(24). https://doi.org/10.3390/molecules27249009 PMid: 36558146 PMCid: PMC9782831

- Sundarraj K, Raghunath A, Perumal E. A review on the chemotherapeutic potential of fisetin: *In vitro* evidence. Biomed Pharmacother. 2018; 97:928-40. https://doi. org/10.1016/j.biopha.2017.10.164 PMid:29136771
- Imran M, Saeed F, Gilani SA, Shariati MA, Imran A, Afzaal M, et al. Fisetin: An anticancer perspective. Food Sci Nutr. 2021; 9(1):3. https://doi.org/10.1002/fsn3.1872 PMid:33473265 PMCid: PMC7802565
- Yan X, Qi M, Li P, Zhan Y, Shao H. Apigenin in cancer therapy: Anti-cancer effects and mechanisms of action. Cell Biosci. 2017; 7(1):50. https://doi.org/10.1186/s13578-017-0179-x PMid:29034071 PMCid: PMC5629766
- Kang CH, Molagoda IMN, Choi YH, Park C, Moon DO, Kim GY. Apigenin promotes TRAIL-mediated apoptosis regardless of ROS generation. Food Chem Toxicol. 2018; 111:623-30. https://doi.org/10.1016/j.fct.2017.12.018 PMid:29247770
- 51. Chen M, Wang X, Zha D, Cai F, Zhang W, He Y, et al. Apigenin potentiates TRAIL therapy of non-small cell lung cancer via upregulating DR4/DR5 expression in a p53-dependent manner. Sci Reports. 2016; 6(1):1-17. https://doi.org/10.1038/srep35468 PMid:27752089 PMCid: PMC5067669
- Rahmani AH, Alsahli MA, Almatroudi A, Almogbel MA, Khan AA, Anwar S, *et al.* The potential role of apigenin in cancer prevention and treatment. Molecules. 2022; 27(18). https://doi.org/10.3390/molecules27186051 PMid: 36144783 PMCid: PMC9505045
- 53. Koirala N, Thuan NH, Ghimire GP, Thang D Van, Sohng JK. Methylation of flavonoids: Chemical structures, bioactivities, progress and perspectives for biotechnological production. Enzyme Microb Technol. 2016; 86:103-16. https://doi.org/10.1016/j.enzmictec.2016.02.003 PMid: 26992799
- Romagnolo DF, Selmin OI. Flavonoids and cancer prevention: A review of the evidence. J Nutr Gerontol Geriatr. 2012; 31(3):206-38. https://doi.org/10.1080/21551 197.2012.702534 PMid:22888839
- 55. Liskova A, Koklesova L, Samec M, Smejkal K, Samuel SM, Varghese E, *et al.* Flavonoids in cancer metastasis. Cancers (Basel). 2020; 12(6):1-29. https://doi.org/10.3390/ cancers12061498 PMid:32521759 PMCid: PMC7352928
- 56. Eltahir S, Ahmad A. Flavonoids on the frontline against cancer metastasis. Cancers (Basel). 2023; 15(16). https:// doi.org/10.3390/cancers15164139 PMid:37627166 PMCid: PMC10452402
- 57. Sahoo S, Mohapatra P, Sahoo SK. Flavonoids for the treatment of breast cancer, present status and future prospective. Anticancer Agents Med Chem. 2023; 23(6):658-75. https://doi.org/10.2174/18715206236662210 24114521 PMid:36284374

- Park MY, Kim Y, Ha SE, Kim HH, Bhosale PB, Abusaliya A, et al. Function and application of flavonoids in breast cancer. Int J Mol Sci. 2022; 23(14). https://doi.org/10.3390/ ijms23147732 PMid:35887080 PMCid: PMC9323071
- 59. Javed Z, Khan K, Herrera-Bravo J, Naeem S, Iqbal MJ, Sadia H, et al. Genistein is a regulator of signalling pathways and microRNAs in different types of cancers. Cancer Cell Int. 2021; 21(1):1-12. https://doi.org/10.1186/s12935-021-02091-8 PMid:34289845 PMCid: PMC8296701
- Raeeszadeh-Sarmazdeh M, Do LD, Hritz BG. Metalloproteinases and their inhibitors: Potential for the development of new therapeutics. Cells. 2020; 9(5). https:// doi.org/10.3390/cells9051313 PMid:32466129 PMCid: PMC7290391
- Niedzwiecki A, Roomi MW, Kalinovsky T, Rath M. Anticancer efficacy of polyphenols and their combinations. Nutrients. 2016; 8(9). https://doi.org/10.3390/nu8090552 PMid:27618095 PMCid: PMC5037537
- 62. Shukla S, Gupta S. Apigenin: A promising molecule for cancer prevention. Pharm Res. 2010; 27(6):962-78. https:// doi.org/10.1007/s11095-010-0089-7 PMid:20306120 PMCid: PMC2874462
- 63. Spagnuolo C, Russo GL, Orhan IE, Habtemariam S, Daglia M, Sureda A, *et al.* Genistein and cancer: current status, challenges, and future directions. Adv Nutr. 2015; 6(4):408-19. https://doi.org/10.3945/an.114.008052 PMid:26178025 PMCid: PMC4496735
- 64. Sharifi-Rad J, Quispe C, Imran M, Rauf A, Nadeem M, Gondal TA, *et al.* Genistein: An integrative overview of its mode of action, pharmacological properties, and health benefits. Oxid Med Cell Longev. 2021; 2021. https:// doi.org/10.1155/2021/3268136 PMid:34336089 PMCid: PMC8315847
- Abotaleb M, Samuel SM, Varghese E, Varghese S, Kubatka P, Liskova A, *et al.* Flavonoids in cancer and apoptosis. Cancers (Basel). 2019; 11(1). https://doi.org/10.3390/ cancers11010028 PMid:30597838 PMCid: PMC6357032
- 66. Zhang HW, Hu JJ, Fu RQ, Liu X, Zhang YH, Li J, *et al.* Flavonoids inhibit cell proliferation and induce apoptosis and autophagy through downregulation of PI3Kγ mediated PI3K/AKT/mTOR/p70S6K/ULK signalling pathway in human breast cancer cells. Sci Rep. 2018; 8(1):11255. https:// doi.org/10.1038/s41598-018-29308-7 PMid:30050147 PMCid: PMC6062549
- Seibold T, Waldenmaier M, Seufferlein T, Eiseler T. Small extracellular vesicles and metastasis-blame the messenger. Cancers. 2021; 13(17):4380. https://doi.org/10.3390/ cancers13174380 PMid:34503190 PMCid: PMC8431296
- Cossarizza A, Gibellini L, Pinti M, Nasi M, Montagna JP, De Biasi S, *et al.* Quercetin and cancer chemoprevention. Evid Based Complement Alternat Med. 2011; 2011. https:// doi.org/10.1093/ecam/neq053 PMid:21792362 PMCid: PMC3136711

- Hosseinzadeh E, Hassanzadeh A, Marofi F, Alivand MR, Solali S. Flavonoid-based cancer therapy: An updated review. Anticancer Agents Med Chem. 2020; 20(12):1398-414. https://doi.org/10.2174/1871520620666200423071759 PMid:32324520
- Gürler SB, Kiraz Y, Baran Y. Flavonoids in cancer therapy: current and future trends. Biodivers Biomed Our Futur. 2020; 403-40. https://doi.org/10.1016/B978-0-12-819541-3.00021-9
- Verdura S, Cuyàs E, Ruiz-Torres V, Micol V, Joven J, Bosch-Barrera J, et al. Lung cancer management with silibinin: A historical and translational perspective. Pharmaceuticals (Basel). 2021; 14(6). https://doi.org/10.3390/ph14060559 PMid:34208282 PMCid: PMC8230811
- Wing Ying Cheung C, Gibbons N, Wayne Johnson D, Lawrence Nicol D. Silibinin--a promising new treatment for cancer. Anticancer Agents Med Chem. 2010; 10(3):186-95. https://doi.org/10.2174/1871520611009030186 PMid: 20015009
- 73. Singh BN, Shankar S, Srivastava RK. Green tea catechin, epigallocatechin-3-gallate (EGCG): Mechanisms, perspectives and clinical applications. Biochem Pharmacol. 2011; 82(12):1807-21. https://doi.org/10.1016/j.bcp.2011. 07.093 PMid:21827739 PMCid: PMC4082721
- 74. Farhan M. Green tea catechins: Nature's way of preventing and treating cancer. Int J Mol Sci. 2022; 23(18). https:// doi.org/10.3390/ijms231810713 PMid:36142616 PMCid: PMC9501439
- 75. Németh K, Plumb GW, Berrin JG, Juge N, Jacob R, Naim HY, *et al.* Deglycosylation by small intestinal epithelial cell β-glucosidases is a critical step in the absorption and metabolism of dietary flavonoid glycosides in humans. Eur J Nutr. 2003; 42(1):29-42. https://doi.org/10.1007/s00394-003-0397-3 PMid:12594539
- 76. Rodriguez-Mateos A, Vauzour D, Krueger CG, Shanmuganayagam D, Reed J, Calani L, *et al.* Bioavailability, bioactivity and impact on the health of dietary flavonoids and related compounds: an update. Arch Toxicol. 2014; 88(10):1803-53. https://doi.org/10.1007/s00204-014-1330-7 PMid:25182418
- 77. Espín JC, Larrosa M, García-Conesa MT, Tomás-Barberán F. Biological significance of urolithins, the gut microbial ellagic acid-derived metabolites: the evidence so far. Evid Based Complement Alternat Med. 2013; 2013. https://doi.org/10.1155/2013/270418 PMid:23781257 PMCid: PMC3679724
- Thilakarathna SH, Vasantha Rupasinghe HP. Flavonoid bioavailability and attempts for bioavailability enhancement. Nutrients. 2013; 5(9):3367. https://doi.org/10.3390/ nu5093367 PMid:23989753 PMCid: PMC3798909
- 79. Zou H, Ye H, Kamaraj R, Zhang T, Zhang J, Pavek P. A review on pharmacological activities and synergistic effect of quercetin with small molecule agents. Phytomedicine. 2021;

92:153736. https://doi.org/10.1016/j.phymed.2021.153736 PMid:34560520

- Kandemir K, Tomas M, McClements DJ, Capanoglu E. Recent advances on the improvement of quercetin bioavailability. Trends Food Sci Technol. 2022; 119:192-200. https://doi.org/10.1016/j.tifs.2021.11.032
- Krautkramer KA, Fan J, Bäckhed F. Gut microbial metabolites as multi-kingdom intermediates. Nat Rev Microbiol. 2020; 19(2):77-94. https://doi.org/10.1038/ s41579-020-0438-4 PMid:32968241
- Iyer N, Corr SC. Gut microbial metabolite-mediated regulation of the intestinal barrier in the pathogenesis of inflammatory bowel disease. Nutrients. 2021; 13(12). https:// doi.org/10.3390/nu13124259 PMid:34959809 PMCid: PMC8704337
- 83. Umamaheswaran G, Krishna Kumar D, Adithan C. Distribution of genetic polymorphisms of genes encoding drug-metabolizing enzymes and drug transporters - A review with an Indian perspective. Indian Journal of Medical Research. 2014; 139:27-65.
- Gummadi AC, Guddati AK. Genetic polymorphisms in pharmaceuticals and chemotherapy. World J Oncol. 2021; 12(5):149. https://doi.org/10.14740/wjon1405 PMid:34804277 PMCid: PMC8577603
- Liu J, Tian M, Wang Z, Xiao F, Huang X, Shan Y. Production of hesperetin from naringenin in an engineered *Escherichia coli* consortium. J Biotechnol. 2022; 347:67-76. https://doi. org/10.1016/j.jbiotec.2022.02.008 PMid:35192875
- Ur Rehman MF, Batool AI, Qadir R, Aslam M. Hesperidin and naringenin. A Centum Valuab Plant Bioact. 2021; 403-44. https://doi.org/10.1016/B978-0-12-822923-1. 00027-3
- Koroleva M, Portnaya I, Mischenko E, Abutbul-Ionita I, Kolik-Shmuel L, Danino D. Solid lipid nanoparticles and nanoemulsions with solid shell: Physical and thermal stability. J Colloid Interface Sci. 2022; 610:61-9. https://doi. org/10.1016/j.jcis.2021.12.010 PMid:34922082
- Sguizzato M, Esposito E, Cortesi R. Lipid-based nanosystems as a tool to overcome skin barrier. Int J Mol Sci. 2021; 22(15):8319. https://doi.org/10.3390/ijms22158319 PMid:34361084 PMCid: PMC8348303
- Wang Z. Knorr quinoline synthesis. Compr Org Name React Reagents. 2010. p. 1638-41. https://doi. org/10.1002/9780470638859.conrr365 PMid:21194205
- 90. Wu B, Basu S, Meng S, Wang X, Zhang S, Hu M. Regioselective sulfation and glucuronidation of phenolics: Insights into the structural basis of conjugation. Curr Drug Metab. 2011; 12(9):900. https://doi.org/10.2174/138920011797470100 PMid:21933112 PMCid: PMC3426368
- Xiao J, Muzashvili TS, Georgiev MI. Advances in the biotechnological glycosylation of valuable flavonoids. Biotechnol Adv. 2014; 32(6):1145-56. https://doi. org/10.1016/j.biotechadv.2014.04.006 PMid:24780153

 Slámová K, Kapešová J, Valentová K. Sweet flavonoids: Glycosidase-catalyzed modifications. Int J Mol Sci. 2018; 19(7). https://doi.org/10.3390/ijms19072126 PMid:30037103 PMCid: PMC6073497

1650

- 93. Soukup ST, Stoll DA, Danylec N, Schoepf A, Kulling SE, Huch M. Metabolism of daidzein and genistein by gut bacteria of the class coriobacteriia. Foods. 2021; 10(11). https://doi.org/10.3390/foods10112741 PMid:34829025 PMCid: PMC8618169
- 94. Yang G, Ge S, Singh R, Basu S, Shatzer K, Zen M, et al. Glucuronidation: Driving factors and their impact on glucuronide disposition. Drug Metab Rev. 2017; 49(2):105. https://doi.org/10.1080/03602532.2017.129368 2 PMid:28266877 PMCid: PMC7660525
- 95. Bondonno NP, Dalgaard F, Kyrø C, Murray K, Bondonno CP, Lewis JR, *et al.* Flavonoid intake is associated with lower mortality in the Danish diet cancer and health cohort. Nat Commun. 2019; 10(1):1-10. https://doi.org/10.1038/s41467-019-11622-x PMid:31409784 PMCid: PMC6692395
- 96. Mazidi M, Katsiki N, Banach M. A greater flavonoid intake is associated with lower total and cause-specific mortality: A meta-analysis of cohort studies. Nutrients. 2020; 12(8): 1-14. https://doi.org/10.3390/nu12082350 PMid:32781562 PMCid: PMC7469069
- Rodríguez-García C, Sánchez-Quesada C, Gaforio JJ, Gaforio JJ. Dietary flavonoids as cancer chemopreventive agents: An updated review of human studies. Antioxidants. 2019; 8(5). https://doi.org/10.3390/antiox8050137 PMid: 31109072 PMCid: PMC6562590
- 98. Arem H, Bobe G, Sampson J, Subar AF, Park Y, Risch H, et al. Flavonoid intake and risk of pancreatic cancer in the National Institutes of Health-AARP Diet and Health Study Cohort. Br J Cancer. 2013; 108(5):1168. https:// doi.org/10.1038/bjc.2012.584 PMid:23299536 PMCid: PMC3619057
- 99. Courtney D, Davey MG, Moloney BM, Barry MK, Sweeney K, McLaughlin RP, et al. Breast cancer recurrence: factors impacting occurrence and survival. Ir J Med Sci. 2022; 191(6):2501. https://doi.org/10.1007/s11845-022-02926-x PMid:35076871 PMCid: PMC9671998
- 100. Arfaoui L. Dietary plant polyphenols: Effects of food processing on their content and bioavailability. Molecules. 2021; 26(10). https://doi.org/10.3390/molecules26102959 PMid:34065743 PMCid: PMC8156030
- 101. Ghosh S, Hazra J, Pal K, Nelson VK, Pal M. Prostate cancer: Therapeutic prospect with herbal medicine. Curr Res Pharmacol Drug Discov. 2021; 2. https://doi.org/10.1016/j. crphar.2021.100034 PMid:34909665 PMCid: PMC8663990
- 102. Lee AH, Su D, Pasalich M, Tang L, Binns CW, Qiu L. Soy and isoflavone intake associated with reduced risk of ovarian cancer in southern Chinese women. Nutr Res. 2014; 34(4):302-7. https://doi.org/10.1016/j.nutres.2014.02.005 PMid:24774066

- 103. Pallmann P, Wan F, Mander AP, Wheeler GM, Yap C, Clive S, *et al.* Designing and evaluating dose-escalation studies made easy: The MoDEsT web app. Clin Trials. 2020; 17(2):147. https://doi.org/10.1177/1740774519890146 PMid:31856600 PMCid: PMC7227124
- 104. Azman M, Sabri AH, Anjani QK, Mustaffa MF, Hamid KA. Intestinal absorption study: Challenges and absorption enhancement strategies in improving oral drug delivery. Pharmaceuticals. 2022; 15(8). https://doi.org/10.3390/ ph15080975 PMid:36015123 PMCid: PMC9412385
- 105. Chen Z, Zheng S, Li L, Jiang H. Metabolism of flavonoids in human: A comprehensive review. Curr Drug Metab. 2014; 15(1):48-61. https://doi.org/10.2174/138920021501 140218125020 PMid:24588554
- 106. Torres-Saavedra PA, Winter KA. An overview of phase II clinical trial designs. Int J Radiat Oncol Biol Phys. 2022; 112(1):22. https://doi.org/10.1016/j.ijrobp.2021.07.1700 PMid:34363901 PMCid: PMC8688307
- 107. Manach C, Scalbert A, Morand C, Rémésy C, Jiménez L. Polyphenols: food sources and bioavailability. Am J Clin Nutr. 2004; 79(5):727-47. https://doi.org/10.1093/ ajcn/79.5.727 PMid:15113710
- 108. Boots AW, Haenen GRMM, Bast A. Health effects of quercetin: from antioxidant to nutraceutical. Eur J Pharmacol. 2008; 585(2-3):325-37. https://doi. org/10.1016/j.ejphar.2008.03.008 PMid:18417116
- 109. Erlund I. Review of the flavonoids quercetin, hesperetin, and naringenin. Dietary sources, bioactivities, bioavailability, and epidemiology. Nutr Res. 2004; 24(10):851-74. https:// doi.org/10.1016/j.nutres.2004.07.005
- Wollenweber E, H. Dietz V. Occurrence and distribution of free flavonoid aglycones in plants. Phytochemistry. 1981; 20(5):869-932. https://doi.org/10.1016/0031-9422(81)83001-4
- 111. Mazzanti G, Menniti-Ippolito F, Moro PA, Cassetti F, Raschetti R, Santuccio C, *et al.* Hepatotoxicity from green tea: a review of the literature and two unpublished cases. Eur J Clin Pharmacol. 2009; 65(4):331-41. https://doi. org/10.1007/s00228-008-0610-7 PMid:19198822
- 112. Du GJ, Zhang Z, Wen XD, Yu C, Calway T, Yuan CS, et al. Epigallocatechin Gallate (EGCG) is the most effective cancer chemopreventive polyphenol in green tea. Nutrients. 2012; 4(11):1679. https://doi.org/10.3390/ nu4111679 PMid:23201840 PMCid: PMC3509513
- 113. Aggarwal BB, Deb L, Prasad S. Curcumin differs from tetrahydrocurcumin for molecular targets, signaling pathways and cellular responses. Molecules. 2014; 20(1):185-205. https://doi.org/10.3390/ molecules20010185 PMid:25547723 PMCid: PMC6272158
- 114. Kay CD. The future of flavonoid research. Br J Nutr. 2010; 104(S3):S91-5. https://doi.org/10.1017/ S000711451000396X PMid:20955652
- 115. Sudhakaran M, Sardesai S, Doseff AI. Flavonoids: New frontier for immuno-regulation and breast cancer

control. Antioxidants. 2019; 8(4). https://doi.org/10.3390/ antiox8040103 PMid:30995775 PMCid: PMC6523469

- 116. Gupta M, Ahmad J, Ahamad J, Kundu S, Goel A, Mishra A. Flavonoids as promising anticancer therapeutics: Contemporary research, nanoantioxidant potential, and future scope. Phytother Res. 2023; 37(11):5159-92. https:// doi.org/10.1002/ptr.7975 PMid:37668281
- 117. Hussain Y, Luqman S, Meena A. Research progress in flavonoids as potential anticancer drugs including synergy with other approaches. Curr Top Med Chem. 2020; 20(20):1791-809. https://doi.org/10.2174/1568026620666 200502005411 PMid:32357817
- 118. Liskova A, Samec M, Koklesova L, Brockmueller A, Zhai K, Abdellatif B, *et al.* Flavonoids as an effective sensitizer for anti-cancer therapy: insights into multi-faceted mechanisms and applicability towards individualized patient profiles. EPMA J. 2021; 12(2):155. https://doi.org/10.1007/s13167-021-00242-5 PMid:34025826 PMCid: PMC8126506
- 119. Wen L, Li G, Huang T, Geng W, Pei H, Yang J, et al. Single-cell technologies: From research to application. Innov Cambridge. 2022; 3(6). https://doi.org/10.1016/j. xinn.2022.100342 PMid:36353677 PMCid: PMC9637996
- 120. YuanD,GuoY,PuF,YangC,XiaoX,DuH,*etal*. Opportunities and challenges in enhancing the bioavailability and bioactivity of dietary flavonoids: A novel delivery system perspective. Food Chem. 2024; 430:137115. https://doi. org/10.1016/j.foodchem.2023.137115 PMid:37566979
- 121. Yetisgin AA, Cetinel S, Zuvin M, Kosar A, Kutlu O. Therapeutic nanoparticles and their targeted delivery applications. Molecules. 2020; 25(9). https://doi.org/10.3390/ molecules25092193 PMid:32397080 PMCid: PMC7248934
- Farhan M, Rizvi A, Aatif M, Ahmad A. Current understanding of flavonoids in cancer therapy and prevention. Metabolites. 2023; 13(4). https://doi.org/10.3390/metabo13040481 PMid: 37110140 PMCid: PMC10142845
- 123. Audulv Å, Hall EOC, Kneck Å, Westergren T, Fegran L, Pedersen MK, *et al.* Qualitative longitudinal research in health research: a method study. BMC Med Res Methodol. 2022; 22(1):1-19. https://doi.org/10.1186/s12874-022-01732-4 PMid:36182899 PMCid: PMC9526289
- 124. Lalu MM, Montroy J, Begley CG, Bubela T, Hunniford V, Ripsman D, *et al.* Identifying and understanding factors that affect the translation of therapies from the laboratory to patients: A study protocol. F1000Research. 2020; 9. https://doi.org/10.12688/f1000research.23663.2 PMid:33123348 PMCid: PMC7570319
- 125. Fernandez-Moure JS. Lost in translation: The gap in scientific advancements and clinical application. Front Bioeng Biotechnol. 2016; 4(JUN):202918. https://doi.org/10.3389/ fbioe.2016.00043 PMid:27376058 PMCid: PMC4891347
- 126. Wagner J, Kroetz D. Transforming translation: Impact of clinical and translational science. Clin Transl Sci. 2016; 9(1):3. https://doi.org/10.1111/cts.12380 PMid:26678255 PMCid: PMC5351317

- 127. Chen L, Cao H, Huang Q, Xiao J, Teng H. Absorption, metabolism and bioavailability of flavonoids: A review. Crit Rev Food Sci Nutr. 2022; 62(28):7730-42. https://doi. org/10.1080/10408398.2021.1917508 PMid:34078189
- 128. Verse F, Janani L, Moradi Y, Solaymani-Dodaran M, Baradaran HR, Rimaz S. Challenges in the design, conduct, analysis, and reporting in randomized clinical trial studies: A systematic review. Med J Islam Repub Iran. 2019; 33(1):37. https://doi.org/10.47176/mjiri.33.37 PMid:31456961 PMCid: PMC6708114
- 129. Butcher NJ, Mew EJ, Monsour A, Chan AW, Moher D, Offringa M. Outcome reporting recommendations for clinical trial protocols and reports: A scoping review. Trials. 2020; 21(1). https://doi.org/10.1186/s13063-020-04440-w PMid:32641085 PMCid: PMC7341657
- Xiong HH, Lin SY, Chen LL, Ouyang KH, Wang WJ. The interaction between flavonoids and intestinal microbes: A review. Foods. 2023; 12(2). https://doi.org/10.3390/ foods12020320 PMid:36673411 PMCid: PMC9857828
- 131. Safe S, Jayaraman A, Chapkin RS, Howard M, Mohankumar K, Shrestha R. Flavonoids: structure-function and mechanisms of action and opportunities for drug development. Toxicol Res. 2021; 37(2):147. https://doi.org/10.1007/s43188-020-00080-z PMid:33868973 PMCid: PMC8007671
- 132. Solnier J, Chang C, Pizzorno J. Consideration for flavonoidcontaining dietary supplements to tackle deficiency and optimize health. Int J Mol Sci. 2023; 24(10). https:// doi.org/10.3390/ijms24108663 PMid:37240008 PMCid: PMC10218363
- 133. Ullah A, Munir S, Badshah SL, Khan N, Ghani L, Poulson BG, et al. Important flavonoids and their role as a therapeutic agent. Molecules. 2020; 25(22). https://doi. org/10.3390/molecules25225243 PMid:33187049 PMCid: PMC7697716
- 134. Redelmeier DA, Zipursky JS. A dose of reality about doseresponse relationships. J Gen Intern Med. 2023; 1-6.
- 135. Kikuchi H, Yuan B, Hu X, Okazaki M. Chemopreventive and anticancer activity of flavonoids and its possibility for clinical use by combining with conventional chemotherapeutic agents. Am J Cancer Res. 2019; 9(8):1517.
- 136. Johnson JL, Adkins D, Chauvin S. A review of the quality indicators of rigour in qualitative research. Am J Pharm Educ. 2020; 84(1):138-46. https://doi.org/10.5688/ ajpe7120 PMid:32292186 PMCid: PMC7055404
- 137. Bentley C, Cressman S, van der Hoek K, Arts K, Dancey J, Peacock S. Conducting clinical trials-costs, impacts, and the value of clinical trials networks: A scoping review. Clin Trials. 2019; 16(2):183-93. https://doi.org/10.1177/1740774518820060 PMid:30628466
- 138. Rist PM, Sesso HD, Manson JAE. Innovation in the design of large-scale hybrid randomized clinical trials. Contemp Clin Trials. 2020; 99:106178. https://doi.org/10.1016/j. cct.2020.106178 PMid:33086158 PMCid: PMC7568770